PURPOSE
This document outlines the standard operating procedures for the Beef Center at the New Deal Farm. Specific experiments or teaching protocols may warrant modifications to the general procedures described below which should be outlined in the approved Animal Care and Use Form authored by the Principle Investigator.

PERSONNEL INVOLVED WITH CATTLE AT THE BEEF CENTER
A. Faculty Coordinator(s) - faculty member(s) officed on campus are responsible for ensuring that the activities at the Beef Center are targeting the mission of the unit; supervisor of Beef Center Manager; must have completed Generic Animal Care and Use Training
B. Principal Investigator - faculty member leading research or teaching activity conducted at the Beef Center or with cattle from the Beef Center; must have completed Generic Animal Care and Use Training
C. Beef Center Manager - full-time staff member officed at the Beef Center are responsible for overseeing all day-to-day management responsibilities; supervisor of part-time student workers; must have completed Generic Animal Care and Use Training
D. Undergraduate or Graduate Student Worker(s) - part-time undergraduate or graduate student workers; must have completed Generic Animal Care and Use Training
E. Teaching or Research Assistants - staff, graduate students, or undergraduate students involved with research projects or teaching activities; supervised by the appropriate Principal Investigator; must have completed Generic Animal Care and Use Training
F. Enrolled Undergraduate or Graduate Students - students interacting with cattle at or from the Beef Center through their respective Animal Science course; supervised by the Principle Investigator of the course or Teaching Assistants listed on the approved Animal Care and Use Form for the respective course; these students are NOT required to have completed Generic Animal Care and Use Training

CLASSES OF CATTLE AT THE BEEF CENTER
A. Breeding Herd - primarily used as a teaching demonstration herd overseen by the Beef Center Manager
   1. Mature Cows (≥24 mos. of age)
   2. Mature Bulls (≥24 mos. of age)
   3. Replacement Heifers (7-24 mos. of age)
   4. Replacement Bulls (7-24 mos. of age)
   5. Heifer Calves (≤ 7 mos. of age)
6. Bull Calves (≤ 7 mos. of age)

B. Custom Developed Cattle - non-TTU owned breeding cattle custom fed for seedstock producers overseen by the Beef Center Manager

1. Bulls (7-24 mos. of age)
2. Heifers (7-24 mos. of age)
3. Experimental Cattle - feedlot or stocker cattle utilized for research overseen by the PI
4. Steers (5-30 mos. of age)
5. Heifers (5-30 mos. of age)
6. Cull cows (≥ 30 mos. of age)

GENERAL PRODUCTION SCHEDULE

A. Breeding Herd

1. Mature Cows
   a. The target is to maintain a ≤ 365 d calving interval.
   b. Cows will be bred during a 60 day breeding season, during which they may be artificially inseminated, naturally serviced, or utilized as a surrogate.
   c. Cows should calve within approximately a 60 day window.
   d. During the calving season, cows will be observed a minimum of twice daily in order to provide assistance if necessary.
   e. Timing of the breeding and calving season will be based on the marketing strategy and/or ability to compliment the current forage system.
   f. Cows not conceiving during the outlined breeding season will be culled.

2. Mature Bulls
   a. Bulls will be administered a Breeding Soundness Exam within 30 to 60 days prior to the initiation of the natural covered breeding season.
   b. Only bulls determined to be a satisfactory breeder should be exposed to cows.
   c. If during the breeding season a stress related event occurs (ie. injury, heat stress, etc.), which may compromise the breeding potential of the bull then he should be re-administered a Breeding Soundness Exam.
   d. At the conclusion of the breeding season, bulls shall be removed from the cows and managed separately until the following breeding season.

3. Replacement Heifers
   a. The target is to have heifers calve for the first time at 20-24 months of age.
   b. Heifers will be managed to target a body weight which is 65% of the estimated mature weight of the female adjusted to Condition Score of 5 at the start of the breeding season.
   c. Heifers will be bred by artificial insemination or natural service.
d. Replacement heifers will be bred 2-4 weeks prior to the initiation of the breeding season for the mature cows.

e. At approximately 12 months of age, heifers will be evaluated for their adjusted 365 day weight, frame score, pelvic size, and pubertal status based upon their reproductive tract score.

f. Carcass ultrasound data may also be collected.

g. In addition heifers will be freeze branded with their permanent identification number on their left hip.

h. Replacement heifers may be sold at any point to other seedstock or commercial operations or kept as replacements for the breeding herd.

4. Replacement Bulls

a. At approximately 12 months of age, bulls will be evaluated for their adjusted 365 day weight, frame score, pelvic size, and administered a Breeding Soundness Exam.

b. Carcass ultrasound data may also be collected.

c. Bulls will be freeze branded with their permanent identification number on their left hip.

d. The vast majority of replacement bulls will be sold at any point to other seedstock or commercial operations.

e. On occasion a yearling bull may be selected for exposure to mature cows or replacement heifers during the breeding season and ownership retained for future use as a mature bull.

5. Heifer Calves

a. Heifer calves are weighed within 24 hours of birth.

b. Simultaneously, an ear tag is placed in the ear demonstrating the dam's identification number, the year code, and the calf number according to the chronological ranking of the calf's birthdate within the herd for that year.

c. The year code plus calf number is then tattooed in one ear. One ear is tattooed at birth and the other at weaning.

d. Calves are weaned at approximately 205 days of age at which point an adjusted 205-day weaning weight is recorded.

e. At weaning, heifer calves may be sold or retained in the replacement heifer group and managed as described above.

6. Bull Calves

a. Bull calves are weighed within 24 hours of birth.

b. An ear tag is placed in the ear demonstrating the dam's identification number, the year code, and the calf number according to the chronological ranking of the calf's birthdate within the herd for that year.

c. The year code plus calf number is then tattooed in one ear. One ear is tattooed at birth and the other at weaning.
d. Calves are weaned at approximately 205 days of age at which point an adjusted 205-day weaning weight is recorded.

e. At weaning, bull calves may be sold or retained in the replacement bull group and managed as described above.

f. If the bull calf is determined not to be herd bull quality either at weaning or prior to weaning they are castrated.

g. In order to minimize stress to the animal, no calves will be castrated beyond 8 months of age. Either surgical or non-surgical castration techniques may be applied.

B. Custom Developed Cattle

1. Bulls

a. Custom developed bulls will be managed similar to the TTU owned replacement bulls described above.

b. An official test period will be setup to measure feedlot performance (ADG, DMI, and G:F) and carcass traits in a given time frame for the customer.

c. Non-virgin bulls will not be allowed on the premise for development.

d. Upon arrival, all bulls are weighed, vaccinated, wormed, and a unique identification tag is placed in the ear if one is not already present.

e. The owner of the cattle will be consulted as to the prior health history and vaccination program of the cattle so that the health program administered at the Beef Center is appropriate.

f. Bulls are kept in dry lots or 10 acre paddocks and provided ad libitum hay and a growing ration in self feeders.

g. After an adequate warm-up period, to allow the cattle to become acclimated to a concentrate diet and remove confounding effects of compensatory gain (approximately 30 days), the bulls are transitioned to a higher concentrate growing ration for an 84 to 114 day test period.

h. Rations fed will either be mixed by the farm manager or provided by a commercial feed company.

i. Initial, interim, and final body weights will be collected.

j. At the end of the testing period, all animals are ultra-sounded by a trained ultrasound technician for intramuscular fat, rib-eye area, back fat thickness at the 12/13th rib interface, and rump fat thickness.

k. Upon completion of the test, bulls will be removed from the premise by the owner(s) of the bulls.

2. Heifers

a. Custom developed heifers will be managed similar to the TTU owned replacement heifers.

b. Procedures outlined for development of heifers will target the objectives of the client including the desired rate of gain, length of time to be developed, variables to be measured, and whether estrus synchronization and artificial insemination is wanted.
3. Upon completion of the development period, heifers will be removed from the premise by the owner(s) of the cattle.

3. Experimental Cattle
   a. Management procedures should be outlined in the approved Animal Care and Use Form for the given experiment.

GENERAL MANAGEMENT PROCEDURES

A. Artificial Insemination
   1. The cow should be restrained in a hydraulic or manual chute.
      a. Restraining the cows head in the head catch of the chute is optional.

   2. The recto-vaginal insemination technique will be utilized.
      a. The technician shall wear a palpation glove on his left hand.
      b. Obstetrical lube should be used to lubricate the anus to provide comfort to the animal based upon the quantity and texture of her manure.
      c. With fingers together in a pointed fashion, the gloved hand is inserted into the rectum and once wrist deep the hand is opened flat against the floor of the rectum such that manure can pass over the top of the hand and arm.
      d. Likewise, the vulva shall be gently wiped with a paper towel to remove excess manure and debris.
      e. The loaded insemination gun should be inserted at a 30° upward angle into the vagina to avoid entering the urethral opening and bladder.
      f. The gun should be raised level once 6 to 8 in inside the vagina.
      g. The vagina and cervix is pulled forward, while gently pushing the gun forward towards the cervix.
      h. Once the gun tip meets the cervix, the external opening of the cervix is gripped with the thumb on top and the forefingers underneath to close the fornix and guide the gun tip into the os cervix.
      i. Once in the cervix, the cervix is manipulated over the gun while gently pushing forward on the gun until with the forefinger the technician can detect the gun tip past the cervix.
      j. Semen is deposited slowly at no more than 1 inch past the cervix into the uterine body.
      k. Once fully dispensed, the gun and the hand in the rectum are removed.

B. Blood Collection
   1. The animal should be restrained in a hydraulic or manual chute and their head restrained in the head catch. The technician should wear latex gloves.
      a. Jugular
         i. A halter should be placed over the animal's head and pulled and tied to the side to further restrain the animal and expose the jugular groove. The jugular vein should be raised by applying pressure at the base of the jugular groove. A needle (18 or 20
(18 or 20 gauge) is then inserted through the hide and into the vessel at an approximately 20\(^\circ\) angle and the blood sample is withdrawn. The needle is removed.

b. Tail
   i. The tail should be gripped with one hand and raised until it is horizontal. The groove located in the ventral midline of the tail should be located approximately 6 in from the base of the tail. A needle (18 or 20 gauge) is inserted perpendicular to the tail midway along the body of a tail vertebrate to draw a blood sample. The needle is removed.

C. Breeding Soundness Exam
   A Breeding Soundness Exam will be administered according to the guidelines outlined by the Society of Theriogenology.

1. Physical traits evaluated without restraint of the animal will be:
   a. Body condition,
   b. Structural soundness,
   c. Eye soundness, and
   d. General overall health.

2. After this portion of the physical examination is completed, the bull should be restrained in a manual or hydraulic squeeze chute.

3. Once restrained, the physical examination can continue with evaluation by palpation of:
   a. The penis for irregularities,
   b. Palpation of the testicles for testicular tone and
   c. Measurement of scrotal circumference.

4. The technician shall wear a palpation glove on his left hand. Obstetrical lube should be used to lubricate the anus to provide comfort to the animal based upon the quantity and texture of his manure.

5. The technician should insert his gloved hand into the rectum and palpate:
   a. The seminal vesicles,
   b. Ampullae, and
   c. Prostate glands for:
      i. inflammation,
      ii. adhesion, or
      iii. fibrosis.
   d. In addition, massaging the accessory sex glands will enhance the sensitivity of the animal for ejaculation.

6. If ejaculation is not achieved through massage, then an electroejaculator will be utilized to encourage an ejaculate sample for seminal evaluation.

7. Prior to insertion of the electroejaculator into the rectum, the gloved hand should be used to clean manure out of the rectum to promote better contact.
8. Once cleaned, the electroejaculator probe will be inserted in the rectum with the electrodes facing ventrally.

9. A hand operated rheostat will be used to provide 2 to 4 second pulses repeated with 5 to 7 second intermissions as the voltage is gradually increased until ejaculation is achieved.
   a. Some bulls may require a quicker pulsation rate to be stimulated.

10. The ejaculate should be collected with a plastic cone attached to a conical vial held within a collection apparatus with a handle.

11. Once the sample is collected, a subsample shall be plated on a slide and viewed microscopically for evaluation of seminal motility and morphology.

12. Bulls shall be classified as satisfactory breeder, unsatisfactory breeder, or deferred for an examination at a later date if the reasons for failing the exam were may change over time.

D. Calving Assistance

Guidelines concerning observation and guidelines for intervention during incidences of dystocia are based upon suggestions by R. G. Mortimer (DVM, Department of Clinical Sciences, Colorado State University).

1. Calving occurs in 3 stages.

2. Stage 1 begins with contraction of the longitudinal and circular muscle fibers of the uterus and ends when the cervix is fully dilated and fetal parts enter the birth canal.
   a. Physical signs of stage 1 include restlessness of the cow, a tendency to lie down, and expulsion and breaking of the chorioallantoic sac (first water sac).

3. Stage 2 begins when the cervix is fully dilated, the amniotic sac (second water sac) is expelled, and the fetus enters the birth canal.
   a. The cow generally lies down and contractions occur 1 to 3 times a minute until the calf is expelled.

4. Stage 3 is expelling of the placenta and fetal membranes post-calving which generally occurs 8 to 12 hours after delivery of the calf.

5. Intervention should occur if the following scenarios occur during each respective stage:
   a. Stage 1
      i. If the cow has been in Stage 1 of labor longer than 8 hours.
   b. Stage 2
      ii. If the water sac is visible for 2 hours and the cow is not trying;
      iii. If the cow has been trying for over 30 minutes and hasn't made any progress;
      iv. If the cow has quit trying for over 20 minutes after a period of progress;
c. Stage 3
   i. If the cow hasn't passed fetal membranes within 24 hours after calving then the cow should be placed in a clean place, the expelled fetal membranes should be regularly disinfected, and the cow should be closely monitored for fever and treated with a long-acting tetracycline as necessary. Placenta should NOT be physically removed.
   ii. Severe cases of retained placenta should be reported to the University Veterinarian or John Hill, DVM in Abernathy, TX for consultation on appropriate treatment for specific cases.

d. Interventions during Stage 1 or 2 of calving may involve restraint of the cow in a head catch or haltering and tying off outside the chute.
   i. If applicable, the technician shall scrub their hands and arm with an antiseptic and clean the vulva.
   ii. If applicable the technician shall use a plastic palpation sleeve. The technician should insert his gloved hand into the vagina and determine the presentation, position, and posture of the calf and decide if they believe they can handle the delivery or if the University Veterinarian or John Hill, DVM in Abernathy, Texas needs to be contacted for a possible cesarean section.
   iii. If the technician believes he can handle the procedure without veterinarian assistance, then obstetrical chains or straps may be used in tandem with a fetal calf extraction device.
   iv. Most importantly, timely decision making by the technician is critical to maximize calf and cow survival during dystocia.

E. Carcass Ultrasound
1. The animal should be restrained in a squeeze chute.
2. An area at least the size of the carcass probe should be shorn to ≤ ¼” horizontally between the 10th rib and 1st lumbar over the top of the longissimus dorsi, vertically between the 12th and 13th rib interface from the thoracic vertebre to the lateral end of the longissimus dorsi, and horizontally over the hook bone.
3. Vegetable oil should be applied liberally to these areas to facilitate contact between the carcass probe and the animal.
4. A 17 cm carcass ultrasound probe should be placed over the three areas described above to capture an image which can be analyzed to estimate the
percentage of intramuscular fat, rib eye area, 12th/13th rib fat depth, and rump fat depth.

F. Dehorning

It is only necessary to use Barnes Dehorner method on calves > 2 months of age (point at which the horn bud attaches to the skull).
1. Restrain the calf appropriately for his size and tie his head to the side using a halter.
2. Use an 18 gauge, 1 ½ in needle, inject 5 ml of banamine (2% Lidocaine) per side 1/3 of the distance from the corner of the eye to the horn bud at the level of the eye. This should be right off the frontal crest bone.
3. Deposit the drug in a fanning action 1 ml at a time for best results by slightly angling the injection in several different directions.
4. Clip the hair around the base of each horn. Set the knife end of the Barnes Dehorner over the horn such that when cutting a ring of hair will be present all the way around the horn (Barnes Dehorners come in different sizes; make sure they are large enough to accommodate the calf).
5. In one motion, aggressively push down and open the dehorners such that they cut underneath the horn.
6. Verify that a circle of hair is present around the horn which you just cut out (if it is not then the horn/scur is likely to grow back); if it is not then re-cut the wound so to remove keratin cells using the Barnes Dehorner or a scoop dehorner. If necessary, pull persistently bleeding arteries with a hemostat.
7. Immediately cauterize the wound. Ensure bleeding has, stopped; however, recognize that burning too long may damage underlying tissues.
8. Apply antiseptic wound spray.
9. Place all surgical tools into an antiseptic wash between calves to reduce incidences of tetanus, bovine cutaneous papillamos virus, and bovine leukosis virus.

G. Castration

The calf will be restrained either in a squeeze chute (larger calves) or by flanking the calf and placing him on his side (smaller calves) and physically holding him down based upon what is determined to be the safest restraint method for the calf and the person performing the castration.
1. Non-surgical
   a. The rubber band will be placed on the elastrator and stretched open in preparation for placement.
   b. The testicles/scrotum will be pulled through the rubber ring and the band dispensed once the technician is confident that ring is placed above the testicles.
   c. The scrotum will be palpated to confirm that neither testicle is above the band and in the abdominal cavity.
   d. An analgesic (ie. banamine) will be administered to help offset chronic pain.
2. Surgical
   a. A local anesthetic (ie. lidocaine) will be administered to larger calves (ie. > 5.5 months of age) via an injection directly into each testicle and then a minimum of 2 minutes should pass prior to initiating castration.
   b. Latex gloves will be worn by the person performing the castration and the scrotum will be disinfected.
   c. The base of the scrotum will be pulled downward with one hand, and with the other hand a scalpel will be used to make a lateral cut one-third of the way down the scrotum.
   d. One hand will be used to palpate and push each testicle outside the scrotum to expose it.
   e. The other hand will be used to grasp the testicles once exposed. Once grasped, each testicle will be pulled away from the body until the spermatic cord is exposed.
      i. It may be necessary to push the scrotal sac upwards to detach it from the testicle.
   f. While grasping one testicle at a time and applying tension, an emasculator will be used to cut and crimp the spermatic cord as high as possible.
      i. The emasculators will be held closed for 5-15 seconds for adequate crimping to help ensure that hemorrhaging does not occur.
   g. An antiseptic powder/spray will be applied to the wound to prevent bacterial infection.
   h. An analgesic (ie. banamine) will be administered to help offset chronic pain. Castration tools will be placed in an antiseptic wash between calves.

H. Embryo Transfer/Flushing
   1. Embryo transfer and flushing techniques should be performed by a professional embryologist (Dr. John Nelson, Nelson Embryo Services, Chickasha, OK).
      a. Donor cows may be flushed on site or at the clinic of the embryologist.
      b. Donor cows will be stimulated with multiple injections of follicle stimulating hormone with dosages recommended by the embryologist.
      c. After synchronization of estrus (as described below), the donor cow will be artificial inseminated by a trained technician (as described above) and 7 day post breeding embryos will be flushed from the uterus and either cryopreserved or transferred fresh into recipient cows.
      d. At the time of flushing, the donor cow will receive an epidural block.
      e. Recipient cows will have estrus synchronized (as described below) and embryos will be transferred at 7 days post estrus.
      f. Recipient cows will receive an epidural block prior to transfer and Banamine afterwards to reduce prostaglandin release and enhance pregnancy rates.
I. Estrus Synchronization
   1. Estrus may be synchronized according to various protocols which are outlined and updated by the Reproductive Beef Task Force. Progesterone (CIDR, MGA, etc.), prostaglandin (Lutalyse, Estrumate, etc), estradiol, and gonadotropin stimulating hormone (ie. Cystorelin, Fertagyl, etc.) maybe utilized to manipulate the timing of ovulation.
   2. Estrus detection aids may be used including tail patches, chalk, electronic devices (Heat Watch System), or a teaser bull with a chin ball marker.
   3. Signs of estrus will be observed a minimum of twice daily in the morning and afternoon.

J. Freeze Branding
   1. Copper branding irons will be placed in liquid nitrogen and allowed to cool for 20 minutes before branding.
   2. Cattle should be restrained in a chute and the squeeze applied.
   3. Cattle will be branded on their left side in their hip area with their permanent identification number and a ranch brand.
      a. This area should be brushed or blown free of dirt and then clipped to less than 1/4 in length.
      b. The area should be saturated with 95-99% isopropyl alcohol using a spray bottle or sponge.
      c. Immediately, afterwards the appropriate brand(s) should be applied to the hide and held firmly.
      d. If surgical clippers were used, the brand should be held to the hide for 25 seconds on calves and 30 seconds on older animals.
      e. If a longer hair guard is used then the brand should be held to the hide for 35 seconds on calves and 40 seconds on older animals.
   4. Branding irons should be cooled for a minimum of 3 minutes between usages.

K. Growth Implant Administration
   1. An appropriate implant administration gun should be utilized relative to the specific implant type being administered.
   2. The needle shall be adequately sharp otherwise it will be replaced.
   3. The needle shall be disinfected between animals.
   4. The technician should administer the implant in the middle 1/3 of the ear in between the rib cartilages.

L. Injections
   1. Procedures involving injections should always follow Beef Quality Assurance standards.
   2. Products should be administered subcutaneously, intramuscularly, intravenously, or intranasally according to the labeled directions.
      a. If possible, subcutaneous administration should be the preferred route of administration over intramuscular.
      b. If intramuscular injection is necessary the injection should be given in the neck.
3. Bent or broken needles should be discarded.
4. Needles will be exchanged at least every 10 animals.
5. The injection needle will never be re-inserted into the bottle.
6. No more than 20 ml of product shall be administered in one location.
7. Withdrawal periods will be noted and cattle will NOT be marketed until they are outside the withdrawal period.

M. Pelvic Size Determination
1. Cattle should be restrained in a chute with a light squeeze.
2. The technician shall wear a palpation glove and use obstetrical lube.
3. Feces should be removed from the rectum.
4. Rice Pelvimeter should be inserted carefully while placed in the palm of the gloved hand.
5. Once inside, the horizontal width of the pelvic inlet at its widest point between the right and left shafts of the ileum should be obtained by squeezing the Rice Pelvimeter and reading the calibration scale on the exterior of the device.
6. Subsequently, the vertical height of the pelvic inlet between the dorsal pubic tubercle on the floor of the pelvis and the sacrum on the top should be obtained by the same means.
7. The horizontal width and vertical width should be multiplied to achieve the pelvic area.
   a. It is recommended that heifers be administered a pelvic area exam 2 to 4 weeks prior to their virgin breeding season so appropriate sire selection decisions can be made relative to calving ease.
   b. Bulls could be evaluated at any time but ideally around 12 months of age.

N. Pregnancy Diagnosis
1. Blood Serum Samples
   a. Blood will be collected as described above.
   b. Samples will be uniquely identified and sent to an appropriate lab for ELISA determination of pregnancy specific protein B.
   c. Pregnancy diagnosis by this means should occur between 28 and 90 days after breeding.
2. Rectal Palpation
   a. The cow should be restrained in a hydraulic or manual chute.
      i. Restraining the cows head in the head catch of the chute is optional.
   b. The technician shall wear a palpation glove on his left hand. Obstetrical lube should be used to provide comfort to the animal based upon the quantity and texture of her manure.
   c. With fingers together in a pointed fashion, the gloved hand is inserted into the rectum and once wrist deep the hand is opened flat against the floor of the rectum such that manure can pass over the top of the hand and arm.
d. The technician then palpates the reproductive tract for uterine tone and size, presence or absence of cotyledons, and/or presence of a palpable fetus or amniotic sac to confirm the pregnancy status.

e. Pregnancy diagnosis by this means should occur between 30 days to term. Earlier term pregnancies should especially be handled with care as not to traumatize the fetus and cause abortion.

3. Ultrasonography
   a. The cow should be restrained in a hydraulic or manual chute.
      i. Restraining the cows' head in the head catch of the chute is optional.
   b. The technician shall wear a palpation glove on his left hand.
   c. Obstetrical lube should be used if the technician to provide comfort to the animal based upon the quantity and texture of her manure.
   d. With fingers together in a pointed fashion, the gloved hand is inserted into the rectum with a 7.5 or 5 MHz reproductive probe in the palm.
   e. The reproductive tract is manipulated with the rectally inserted hand until an acceptable image can be captured of the fetus or amniotic sac (or lack thereof) from which pregnancy status can be confirmed

O. Reproductive Tract Score Assessment
   1. Rectal palpation or ultrasonography of the reproductive tract are both acceptable means to evaluate the pubertal status of a heifer using a reproductive tract scoring system.
      a. The reproductive tract score exam should be administered prior to the initiation of the heifer's virgin breeding season for decision making purposes.
      b. Rectal palpation or ultrasonography procedures would be similar to those described above under pregnancy diagnosis except instead of confirming the pregnancy status the uterine horn size and tone, size of the ovaries, and structures on the ovary are utilized as clues to assess the reproductive tract score on a scale between 1 and 5.

P. Tagging
   1. Older cattle should be restrained in a chute for tagging purpose.
   2. Younger calves may be caught and handheld. Universal or electronic tags and their appropriate applicator may be used to place a uniquely numbered ear tag in the middle 1/3 of the ear between rib cartilages.

Q. Torching/Clipping
   1. Cattle may be clipped, torched, or sheared to enhance their phenotype for marketing or exhibition purposes.
      a. Cattle will be restrained in a chute if necessary or tied with a halter if halter broke.
      b. The technician shall confirm that no flammable objects are around the area or on the animal before torching.
i. Specifically, torching should not be performed directly after an animal as had a topical anthelmintic applied.

c. A propane tank outfitted with a hose and a torch is used for singeing the rough hair off the animal at a distance of 6 to 12 inches depending upon the length of the hair.
   i. Torching should be avoided around the eyes and ears of the animal and should be performed with quick motions as to avoid burning the animal.

d. Clipping may be performed with a variety of sheep head or hair head blades depending upon the desired length and area to be clipped by the technician.

e. In both torching and clipping procedures the technician shall be aware of the animal’s behavior and not put themselves in a dangerous position in which they may be kicked.

GENERAL NUTRITION PROGRAM

A. January 1-March 1
   1. Alfalfa hay, gin trash, haygrazer hay, etc. (forage based diet), plus free choice mineral and Sweeet Bran is used as an energy and protein supplement.

B. March 1-May 1
   1. As above until calving, then cows with small calves are moved to wheat pastures with high Mg free choice mineral.

C. May 1-July 1
   1. Alfalfa, wheat, sorghum, or sorghum-sudan, and free choice mineral (grain sorghum or corn may be used as energy supplement if needed)

D. July 1-October 1
   1. Grazing pasture or hay as available.

E. October 1-January 1
   1. Crop stubble fodder or hay as available.

F. Formulated Ration
   1. The PP3 ration is mixed and delivered by the farm’s feed mill.
   2. This ration is primarily used for development of heifers and young bulls.
   3. Ingredient composition is approximately as listed below unless fluctuations in ingredient costs warrant modification:
      
      | Ingredient              | Percentage |
      |-------------------------|------------|
      | Cotton Seed Hulls       | 29.5%      |
      | Corn                    | 48.4%      |
      | Soybean Meal            | 13.3%      |
      | Salt                    | 1.8%       |
      | CaCO₃                   | 0.9%       |
      | Vitamin A Premix        | 0.3%       |
      | Molasses                | 5.8%       |

ANIMAL HEALTH PROGRAM
A. Records
   1. All routine health care is performed by the Beef Center Manager.
   2. Herd health and observation records are maintained and kept on file in the Beef Center office.
   3. All animals are observed at least once daily for general health and performance assessment.

B. General Considerations
   1. Animal health products requiring a prescription are purchased through the University Veterinarian or through a local veterinarian.
   2. All other non-prescription pharmaceuticals are purchased through animal health product distributors.
   3. All serious and non-routine illnesses are brought to the attention of a veterinarian for proper diagnosis and treatment.
   4. In addition to the University Veterinarian, John Hill, D.V.M. in Abernathy, Texas may be consulted for diagnosis and treatment of sick cattle and/or Mike Mimms, D.V.M. in Hereford, TX.

C. Mature Cows and Bulls
   1. All cows and bulls are re-immunized annually with
   2. Modified live vaccine to protect against:
      a. Bovine respiratory disease (IBR, PI3, BVD Types I and II, BSRV),
      b. Pasteurella and Haemophilus somnus,
      c. Leptospirosis (L. pomona, L. canicola, L. icterohemorrhagica, L. grippotyphosa, and L. borgpetersenii serovar hardjo bovis) and
      d. Campylobacteriosis fetus
   3. Vaccinations are given prior to breeding according to the manufacturer's directions.
   4. Cows and bulls are given an annual booster vaccination against the clostridial diseases (Cl. chauvei, Cl. septicum, Cl. Novyi Type B & D, Cl. sordelli, and Cl. perfringens C & D) with a seven-or eight-way bacterin toxoid.
   5. All adult cattle are treated with a systemic insecticide to control grubs each summer, and are sprayed as needed to control lice.
   6. All adult cattle are treated twice per year (spring and fall) with an anthelmintic to control internal and external parasites.

D. Replacement Heifers and Bulls
   1. Heifers should be vaccinated for:
      a. Brucellosis between 4 and 10 months of age by John Hill, DVM in Abernathy, TX
      b. Other immunization procedures should follow that as described with adult cattle.
   2. Bulls which are sold may require PCR testing for Trichomoniasis foetus prior to being marketed by John Hill, DVM in Abernathy, Texas.

E. Calves
1. All calves are to be immunized at approximately three and six months of age with:
   a. A modified live vaccine to protect against:
      i. Bovine respiratory disease (IBR, PI3, BVD Types I and II, BSRV),
      ii. Pasteurella and Haemophilus somnus,
      iii. Leptospirosis (L. pomona, L. canicola, L. icteromhemorrhagica, L. grippotyphosa, and L. borgpetersenii serovar hardjo bovis) and
      iv. Campylobacteriosis fetus.
   b. Vaccinated and re-vaccinated against the clostridial diseases (Cl. Chauvei, Cl. septicum, Cl. Novyi Type B & D, Cl. sordellii, and Cl. perfringens C & D) with a seven-or-eight-way bacterin toxoid.

F. Experimental Cattle
   1. Exact health processing procedures should be outlined in the approved Animal Care and Use Form for the given experiment.
   2. Light weight cattle (250 to 600 lb) that are brought in for research purposes are:
      a. Given vaccinations for Clostridial and bovine respiratory diseases upon arrival,
      b. Booster injections 7 to 21 days later, depending upon source of cattle.
      c. Antibiotics may be administered depending upon source of cattle. Animals are also treated with a commercial parasite control product (may be injectable, pour-on or drench).
      d. These cattle may also receive a growth promoting implant at or shortly after arrival.
      e. These animals will receive an ear tag to uniquely identify them.
   3. Heavier cattle (700 to 900 lb at arrival) may receive these same treatments if the background is not known, but may not if these practices have already occurred before shipment to the Beef Center.

G. Identification and Treatment of Infectious Diseases
   1. Many cattle health problems at the Beef Center are considered to be routine.
   2. Symptoms shall be noted and treatment shall be applied based upon the expertise of the Beef Center Manager.
   3. If the Beef Center Manager is uncertain as to the diagnosis or upon the proper treatment the University Veterinarian shall be consulted.
   4. In rare instances, extremely ill animals may need to be euthanized.
      a. If this is necessary, cattle may be euthanized by gunshot to the head or lethal injection under veterinary recommendations.

H. Animal Disposal
   1. Dead animals should be immediately removed from the pen or paddock and disposed of in the landfill located in the Southeast paddock of the Beef Center.

FACILITIES
   A. Paddocks
1. Approximately 650 acres of the New Deal facility are utilized in support of the Beef Center.
   a. The Beef Center Manager is responsible for maintaining fences and managing these pastures which may include weed control, fertilization, annual forage establishment, and/or permanent forage establishment and haying.
   b. Annual forages which may be planted for grazing or haying purposes include sorghum-sudan, wheat, and oats.
   c. All pastures have their own water troughs.

B. Holding Pens
1. Two holding pens with cover adjacent to animal working facilities are provided for sick animals.
2. In addition there are two more small holding pens next to the load out area.
3. There are nine larger pens for housing of cattle for teaching, research, or maintenance objectives.
4. All pens have water troughs and feed bunks.

C. Research Pens
1. There are 48 research pens located on the Southwest corner of the Beef Center's dry-lot area.
   a. Each pen is 30 ft x 10 ft in size and contains a concrete apron, concrete feed bunk, and individual water trough.
   b. A platform scale is located on the west end to facilitate group weighing of pens of experimental cattle at interim periods.

D. Office and Working Chute
1. The Beef Center Manager's office is located on the Northwest corner of the dry-lot area.
2. Records shall be kept and maintained in the office.
3. A refrigerator is used to maintain pharmaceutical products requiring refrigeration.
4. Adjacent to the office is a livestock tub which leads to a long alley and hydraulic chute containing weight scales where cattle can be restrained.

E. Animal Science Livestock Arena
1. On occasion, cattle from the Beef Center may be transported to the Animal Science Livestock arena located on campus for teaching purposes.

TRANSPORTATION AND HANDLING OF LIVESTOCK
A. Cattle Handling
1. Care shall be taken not to provoke stress or excitement of cattle when being shipped or processed. Electronic prods and whips should only be used as last resort to encourage cattle movement.

B. Cattle Transportation
1. Cattle shall be hauled in a 7 ft x 28 ft or 7 ft x 24 ft gooseneck trailer and truck owned by the Animal and Food Sciences Department. Refer to SOP030 Animal Transportation.

FACILITY MAINTENANCE

A. The herd manager takes care of all routine maintenance, which includes:
   1. Electrical, irrigation, building, and regular vehicle maintenance.
   2. If a situation occurs when the herd manager is not able to repair something, then independent service personnel will be called at the manager’s discretion and the service personnel’s availability.
   3. Texas Tech University Building Maintenance and Utilities is called if the herd manager cannot make repairs.

LEASING OF CATTLE

A. The Faculty Coordinator reserves the right to lease cattle to other entities including but not limited to:
   1. Other academic institutions,
   2. Livestock organizations, and
   3. Individual cattle producers as it targets the teaching and outreach mission of the department.
B. Conditions of the lease shall be outlined in a separate document and signed by both parties.
   1. Conditions outlined in the agreement should include but not be limited to the duration and timing of the lease, the compensation for leasing the animal(s), and the party responsible for taking care of the animal(s) during the lease.
   2. During the time that animal(s) are engaged in the lease and not at the Beef Center they shall not be required to be managed according to the Beef Center SOP022 or be required to be managed underneath an approved IACUC protocol until they are returned to the Beef Center premise.