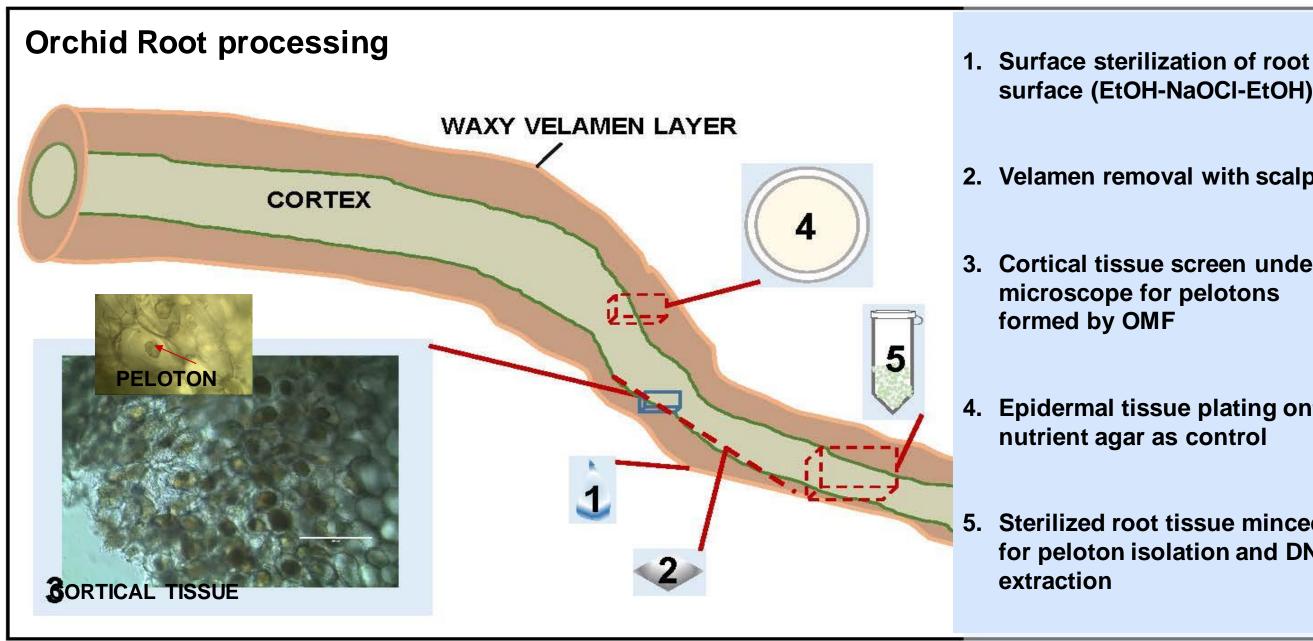


# Endofungal bacteria isolated from mycorrhizal fungi in a North American terrestrial orchid Kris Petterson, Jaspreet Kaur, and Jyotsna Sharma **Department of Plant and Soil Science, Texas Tech University**

### Introduction

Orchidaceae is among the largest and most diverse plant families consisting of approximately 30,000 orchid species worldwide with the majority of these being rare in nature because of their specialized ecological niches. To gain a mechanistic understanding of their complex ecological interactions, and to inform *in-situ* conservation, it is important to first discover and describe the diversity of organisms that orchids associate with. All orchids form obligate mycorrhizae with orchid mycorrhizal fungi (OMF), and accordingly, OMF diversity and interactions are commonly studied. However, endophytic bacteria are also known to enhance biomass and survival of orchid seedlings in vitro, suggesting another important symbiotic niche for orchids. Yet, orchid associated bacteria remain understudied, especially with respect to their role as mycorrhizal helper bacteria (MHB). We report here the first results of our investigation of culturable Endophytic Fungi (EF) and Endophytic Bacteria (EB), with specific focus on OMF and Endofungal Bacteria (EFB). We selected a North American terrestrial orchid, *Platanthera* chapmanii, for our studies, and sampled its roots across two years from two disjunct occurrences in Texas and Florida. Cultured fungal and bacterial isolates were identified by amplifying and sequencing the nrITS and 16S regions, respectively. We also ascertained the endosymbiotic status of bacteria within fungal hyphae by using scanning electron microscopy (SEM). To the best of our knowledge, ours is the first report of isolation of endofungal bacteria from orchid mycorrhizal fungi.

### Methods



#### Symbionts targeted for culture-based isolation

Peloton-forming OMF and their EFB (EF and EB were simultaneously cultured)

#### **Isolation Media**

OMF and EF – Modified Melin-Norkrans (MMN) and Potato Dextrose Agar (PDA) EB – Czapek Agar (CZ) and King's B Media (KB)

#### Endofungal bacterial isolation

PDA with Ampicillin with Streptomycin Malt Extract Agar (MEA) with Cycloheximide

#### Visualization of hyphae

Scanning electron microscopy (SEM) of EF and OMF from above

#### **DNA-based identification of fungi and bacteria**

Amplification and sequencing of fungal nuclear ribosomal ITS region with primer pairs ITS1/ITS4OF and ITS1/ITS4-Tul

Amplification of the bacterial 16S region with primer pairs 27F/1492R and 515F/916R



#### **Research Questions**

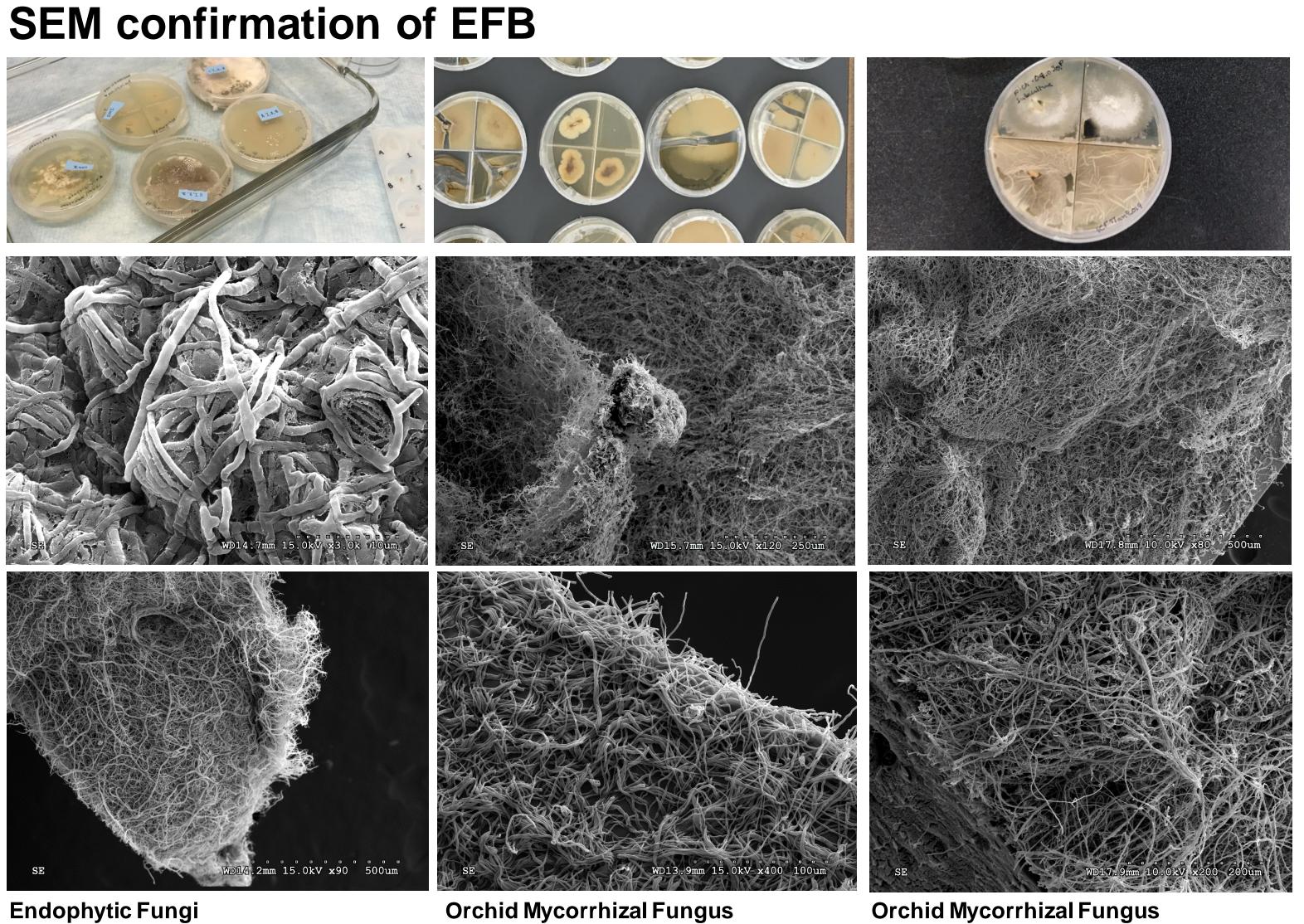
. Are endofungal bacteria present and culturable from endophytic or orchid mycorrhizal fungi?

2. Do plants of *Platanthera chapmanii* from widely separated, disjunct populations host distinct orchid mycorrhizal fungal and / or endofungal bacterial communities?



- . Velamen removal with scalpel
- Cortical tissue screen under
- . Epidermal tissue plating onto
- Sterilized root tissue minced for peloton isolation and DNA





plch.04.05.01P Fusarium coffeatum

plch.15.02.02P **Uncultured Ceratobasidiaceae** 

Endofungal bacteria [b.39.01(plch.15.02.01 on MEA)] Bacillus subtilis strain 1-17

plch.04.03.01T Uncultured Tulasnellaceae

Endofungal bacteria [b.31.01(plch.04.03.01T on MEA)] Bacillus subtilis

## Texas (2018) 12 unique OMF OTUs 6 unique EF OTUs 6 unique EB OTUs 1 unique EFB OTU

### Bacteria; OTU – Operational Taxonomic Unit

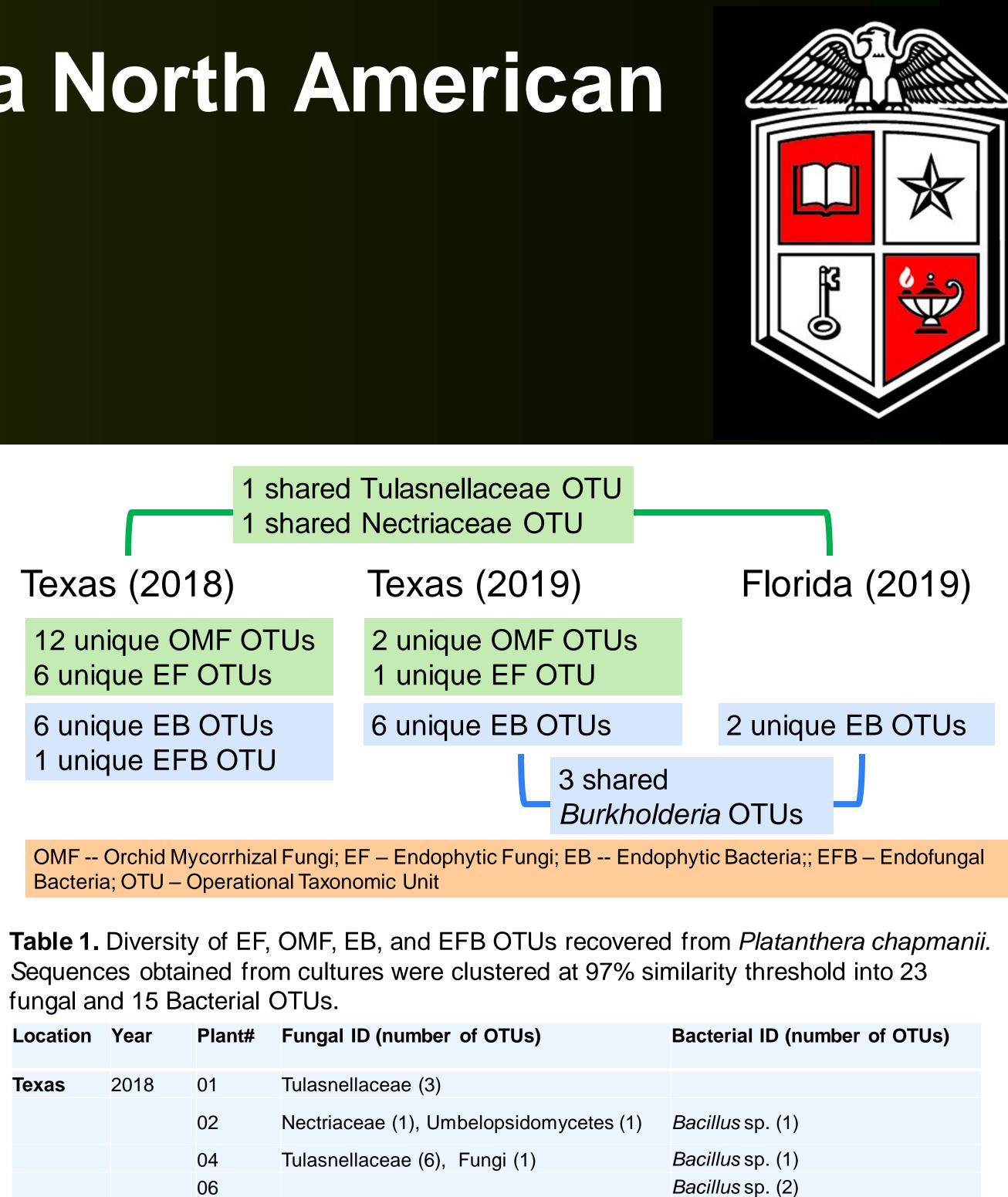
fungal and 15 Bacterial OTUs. Location Year

Texas	2018	01
		02
		04
		06
		07
		08
		10
		11
		12
		13
		14
		15
	2019	03
		04
		05
		07
		10
		11
		12
		14
		15
		16
Florida	2019	01
		03
		04
		05
		10
		11
		13
		14

#### Summary and Conclusions

#### Acknowledgements

We gratefully acknowledge research funding or other support from: Texas Parks and Wildlife Department, Watson Native Plants Preserve, Big Thicket National Preserve, and the Conservation Committee of the Southwest Regional Orchid Growers Association (SWROGA). We thank Joe Liggio (Houston, TX), Houston Snead (Jacksonville Zoo, Jacksonville, FL), and Shan Wong for their assistance with activities in the field.



Tulasnellaceae (1) Tulasnellaceae (1)

Fungi (1) Ascomycota (1) Nectriaceae (1), Hypocrealles (1) Ceratobasidiaceae (1)

Umbelopsidomycetes (1) Sodariomycetes (1) Ceratobasidiaceae (1)

Ceratobasidiaceae (1)

Paenibacillus sp. (1) Burkholderia sp. (1) Bacillus sp. (1), Burkholderia sp.(1) Burkholderia sp. (1) Burkholderia sp. (3)

Burkholderia sp. (1), Bacteria (1)

Burkholderia sp. (1), Bacteria (1)

Burkholderia sp. (1)

Burkholderia sp. (1)

Burkholderia sp. (1)

Bacillus sp. (1)

Bacillus sp. (1)

Bacillus sp. (1)

Tulasnellaceae (1)

Nectriaceae (1)

• Our study is the first to isolate endofungal bacteria from orchid mycorrhizal fungi (OMF).

• In this ongoing work, we have thus far obtained 95 individual fungal cultures (EF and OMF combined), 105 individual EB cultures, and isolated and cultured 21 EFB. Of these, acceptable amplification and sequencing (>200 bp) is complete for 15 EF, 23 OMF, 25 EB, and 16 EFB.

• Altogether, we recovered 16 isolates of endofungal bacteria (EFB) from OMF (6) and EF (10). Taxonomically, the 16 isolates of EFB clustered into 1 Burkholderia OTU and 2 Bacillus OTUs. The 23 OMF isolates clustered into 12 Tulasnellaceae OTUs and 2 Ceratobasidiaceae OTUs.

• Further, the 15 EF isolates clustered into 3 unspecified fungal OTUs, 1 Hypocrealles OTU, 2 Nectriaceae OTUs, 1 Ascomycota OTU, 1 Umbelopsidomycetes OTU, and 1 Sodariomycetes OTU. The 25 EB isolates were clustered into 7 Burkholderia OTUs, 2 Bacillus OTUs, 1 Paenibacillus OTU, and 2 unspecified bacterial OTUs.

• Our research will ultimately steer an understanding of specialized, tripartite interactions of OMF, their endofungal bacterial symbionts, and orchids.